

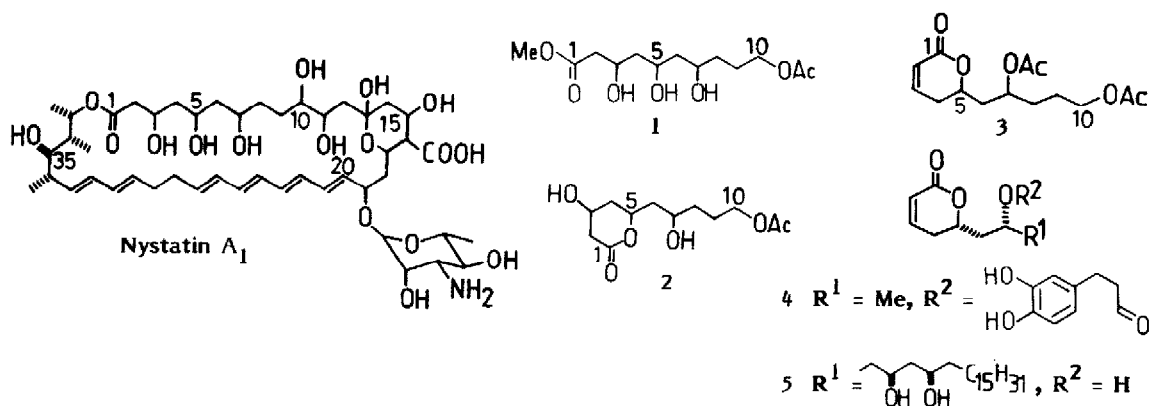
STEREOSTRUCTURE OF NYSTATIN A₁: A SYNTHETIC ASSIGNMENT OF THE C1-C10 FRAGMENT

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Summary: The 3*R*, 5*R* and 7*R* configurations for nystatin A₁ have been assigned through a comparison of synthetic and natural fragment 3. The key step of the synthetic sequence is a nucleophilic oxirane opening by an α -glycosyl copper reagent.

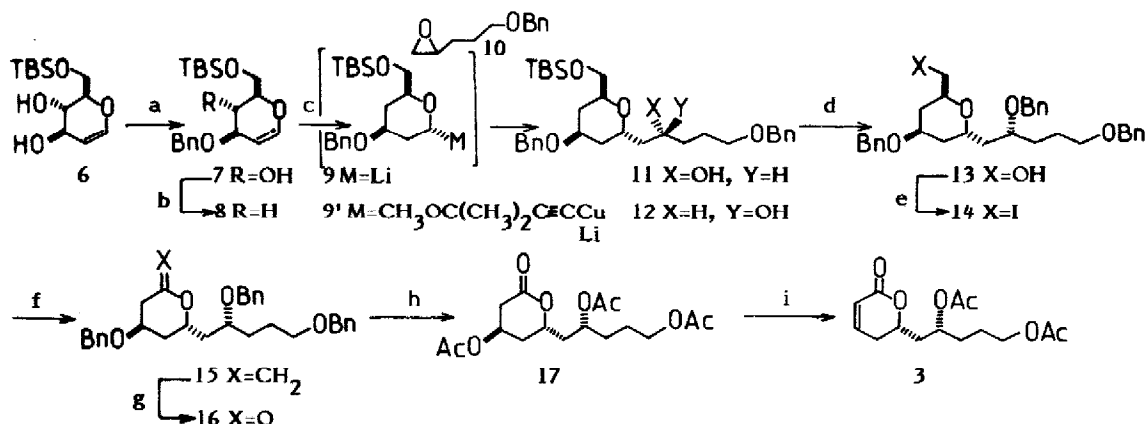
We recently reported¹ the 3*R**, 5*R** and 7*R** relative configurations for nystatin A₁, a commonly used antibiotic in human therapy². The stereochemical information was deduced by detailed proton-nmr analyses of the two degradation C1-C10 fragments, 1 and 2. Exhaustive acetylation (Ac₂O, pyridine) and purification led to the unsaturated δ -lactone 3, $[\alpha]_D^{20}$ -62° (c 0.23, CHCl₃).



In order to determine the absolute configuration of this molecular segment of nystatin A₁, we undertook the synthesis of either 1, 2 or 3. Of the two possible enantiomeric candidates, the 3*R*,5*R*,7*R* compound was considered to be the most likely target for the synthetic work, based on the structural resemblance of nystatin A₁ to amphotericin B³, and on the optical rotation of 3, closely related to that of 4⁴, $[\alpha]_D$ -67° (c 2.3, CHCl₃), and 5⁵, $[\alpha]_D$ -34° (MeOH), the α,β -unsaturated δ -lactones of plant origin whose stereochemistry was recently established⁶.

In continuation with our work on anionic anomeric species in the 2-deoxy series⁷, we became interested in the possibility of expanding further their use in the synthesis of the 1,3-polyol systems. Within this context, an obvious strategy for preparing a C1-C10 nystatin A₁ segment which contains the required chiral centers would be to assemble a sugar-derived anionic compound (C1-C5)⁸ with a suitable five-carbon epoxide (C6-C10)⁸. For the stereoselective carbon-carbon bond formation, we capitalized on the fact that the 2-deoxy glycosyl lithium⁷ and copper⁹ reagents are configurationally stable at low temperatures.

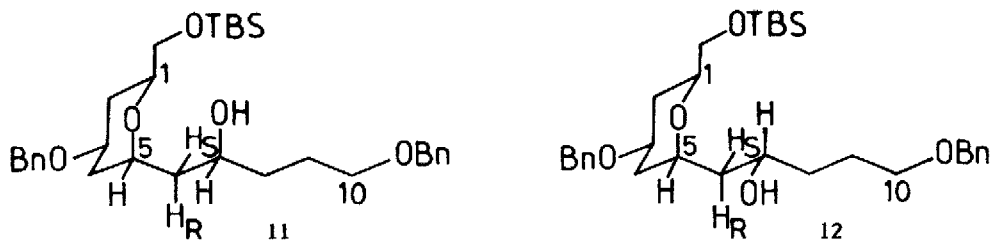
The known 6-O-silylated-D-glucal¹⁰, available in two steps from tri-O-acetyl-D-glucal, was regioselectively benzylated at the 3-O-position via the stannylene procedure¹¹ to give the 3-O-



Scheme: (TBS = *t*-butyldimethylsilyl) (a) Bu₂SnO, NBu₄Br, BnBr, PhCH₃, 80°C, 18 h, 93%; (b) NaH, CS₂, r.t., 4 h then CH₃I, r.t., 18 h; Bu₃SnH, cat. AIBN, PhCH₃, reflux, 1.5 h, 73%; (c) HCl, PhCH₃, 1 min, -15°C then LN, THF, -78°C; CH₃OC(CH₃)₂C≡CCu, THF, -78°C then 10, BF₃:Et₂O, 5 equiv., -78 to 0°C, 1 h, 36%; (d) NaH, BnBr, DMF, 0°C, 1 h; NBu₄F, THF, r.t., 2 h, 86%; (e) Ph₃P, imidazole, I₂, PhCH₃, r.t., 3 h, 68%; (f) DBU, THF, reflux, 15 h; (g) O₃, CH₂Cl₂, 1 min, -78°C then Me₂S, 75% from 14; (h) H₂, Pd/C, AcOEt, r.t. then Ac₂O, pyridine, r.t. quant.; (i) DBU, PhCH₃, r.t., 15 min.

benzylated glucal 7¹² (93% yield, Scheme). Subsequent reductive removal of the C3 hydroxyl group by the xanthate protocol¹³ provided the disymmetrically protected glycal 8 (73% yield), $[\alpha]_D^{25} -18^\circ$ (c 0.86, CHCl₃) recently described by Paquette¹⁴. The unstable α -D-glycopyranosyl lithium derivative 9, prepared by hydrochlorination in toluene⁷ and reductive lithiation with lithium naphthalenide in THF at -78°C, was converted into the presumed mixed organocuprate reagent 9'. Reaction of a THF solution of 9' with the racemic epoxide 10, in the presence of the BF₃:OEt₂ complex¹⁶ (5 equiv.) at -78°C led to the diastereoisomeric alcohols 11 and 12¹⁷ (36% yield from glycal 8). Despite the moderate yield¹⁸ of this reaction, two interesting features are worth mentioning. The α -C-glycosidic products were the only coupling products formed, without any of the β -compounds being detected. This was confirmed by oxidation of alcohols 11 and 12 which led to a single ketone¹⁹. The known⁷ configurational stability of the initial lithiated species is thus preserved during the course of the following anionic modifications. A noticeable degree, although modest (11:12 ratio, 3:2), of stereoselectivity in the desired sense was observed by the reaction of the chiral anionic species with the racemic epoxide. The kinetic preference may well become synthetically useful with an appropriate structural modification of the reactants.

The configuration at C7 (nystatin numbering) was unambiguously established by proton-nmr analysis of 11 and 12 in CDCl₃¹⁹ (or C₆D₆) solution. The values of the 3J and 4J coupling constants demonstrate that the tetrahydropyran ring adopts a chair conformation. The assignment of the coupling constants for H_R at C6 ($^3J_{6R,7}$ 2.2, $^3J_{5,6R}$ 2.5 and $^4J_{6R,6S}$ 14.8 Hz in 11; $^3J_{5,6R}$ 3.8, $^3J_{6R,7}$ 8.4 and $^4J_{6R,6S}$ 14.5 Hz in 12) and H_S at C6 ($^3J_{6S,7}$ 9.4, $^3J_{5,6S}$ 11.4 and $^4J_{6R,6S}$ 14.8 Hz in 11; $^3J_{6S,7}$ 3.1, $^3J_{5,6S}$ 10.8 and $^4J_{6R,6S}$ 14.5 Hz in 12) is only compatible with the configuration shown (see below) where the C6-C10 appendage appears to take an extended zig-zag conformation, or a slight deviation from it in compound 11. In the latter case, an intramolecular hydrogen bonding between O5 and the hydroxyl group at C7 stabilizes the conformation observed as already detected in this type of situation¹. The unwanted isomer was transformed (Ph₃P, DEAD, PhCOOH, THF, r.t.) to the one desired using the Mitsunobu reaction²⁰.



Benzylation of 11 and subsequent desilylation with NBu_4F in THF afforded the primary alcohol 13 converted to iodide 14 using the standard methodology²¹. DBU in THF easily eliminated hydrogen iodide from 14 and the unsaturated compound 15 was immediately ozonolyzed to give lactone 16, $[\alpha]_{\text{D}}^{20} +3^\circ$ (c 0.65, CHCl_3) after reductive workup. Lactone 16 has the full structure and substitution pattern of the C1-C10 nystatin A_1 segment.

For purposes of comparison with degradation product 3, benzyl ethers were hydrogenolyzed in ethyl acetate and the resulting triol acetylated. This sensitive triacetate 17, contaminated by a small amount of α,β -unsaturated lactone 3, was readily converted into 3 by a brief treatment with DBU in toluene. Synthetic 3²² showed identical chromatographic and spectroscopic data and the same optical rotation, $[\alpha]_{\text{D}} -64^\circ$ (c 0.11, CHCl_3) as compound 3 derived from nystatin A_1 .

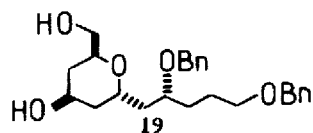
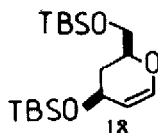
In conjunction with previous studies¹, the 3R, 5R and 7R configurations for the antibiotic nystatin A_1 are thus established²³. Complete stereostructure of nystatin A_1 is described in the following communication.

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 17. Purification of the coupling compounds **11** and **12** was greatly facilitated by acetylation of the crude product and isolation of the obtained acetates. Chromatography of the deacetylated mixture afforded alcohols **11** [$\alpha_D^{20} +19^\circ$ (c 1.6, CHCl_3), and **12** [$\alpha_D^{20} +21^\circ$ (c 1.17, CHCl_3).
 18. The disilylated glycol **18** gave much better results. Selective displacement at the primary position of diol **19** (e.g., step e, in Scheme) under a variety of conditions was however unmanageable in our hands due to a competitive displacement of the hydroxyl at position 3 (glycol numbering) and/or the 3,6-anhydro ring formation.



19. Selected ^1H -nmr data (nystatin A_1 numbering is used): **11** (CDCl_3): δ 1.36 (m, $J_{6R,7} = 2.2$, $J_{5,6R} = 2.5$, $J_{6R,6S} = 14.8$ Hz, H6R); 1.44 (dt, $J_{1,2a} = J_{2a,3} = 8.5$, $J_{2a,2e} = 13.1$ Hz, H2a); 1.85 (ddd, $J_{6S,7} = 9.4$, $J_{5,6S} = 11.4$, $J_{6R,6S} = 14.8$ Hz, H6S); 1.94 (m, $J_{2e,4e} \sim 1$, $J_{1,2e} = J_{2e,3} = 3.9$; $J_{2a,2e} = 13.1$ Hz, H2e); 4.26 (m, $J_{5,6R} = 2.5$, $J_{4,5} = 4.5$, $J_{4',5} \sim 5$, $J_{5,6S} = 11.4$ Hz, H5). **12** (CDCl_3): δ 1.39 (ddd, $J_{5,6R} = 3.8$, $J_{6R,7} = 8.4$, $J_{6R,6S} = 14.5$ Hz, H6R); 1.44 (dt, $J_{1,2a} = J_{2a,3} = 8.0$; $J_{2a,2e} = 13.5$ Hz, H2a); 1.89 (ddd, $J_{6S,7} = 3.1$, $J_{5,6S} = 10.8$, $J_{6S,6R} = 14.5$ Hz, H6S); 1.96 (dt, $J_{1,2e} = J_{2e,3} = 3.9$, $J_{2a,2e} = 13.5$ Hz, H2e); 4.37 (m, $J_{5,6R} = 3.8$, $J_{4,5} \sim 5$, $J_{4',5} \sim 6$, $J_{5,6S} = 10.8$ Hz, H5). Ketone derived from **11**, **12** (C_6D_6): δ 1.58 (m, $J_{2e,4e} = 1.7$, $J_{3,4e} = J_{4e,5} = 4.3$, $J_{4a,4e} = 13.3$ Hz, H4e); 1.70 (ddd, $J_{4a,5} = 4.9$, $J_{3,4a} = 8.7$, $J_{4a,4e} = 13.3$ Hz, H4a); 2.03 (dd, $J_{5,6S} = 6.0$, $J_{6R,6S} = 15.2$ Hz, H6S); 2.51 (dd, $J_{5,6R} = 8.2$, $J_{6R,6S} = 15.2$ Hz, H6R); 4.55 (m, $J_{4e,5} = 4.3$, $J_{4a,5} = 4.9$, $J_{5,6S} = 6.0$, $J_{5,6R} = 8.2$ Hz, H5). **16** (CDCl_3): δ 1.68 (ddd, $J_{3,4a} = 3.8$, $J_{4a,5} = 11.7$, $J_{4a,4e} = 13.7$ Hz, H4a); 2.11 (m, $J_{2e,4e} = 1.7$, $J_{4e,5} = 2.9$, $J_{3,4e} = 3.8$, $J_{4a,4e} = 13.7$ Hz, H4e); 2.64 (dd, $J_{2a,3} = 5.2$, $J_{2a,2e} = 12.5$ Hz, H2a); 2.75 (ddd, $J_{2e,4e} = 1.7$, $J_{2e,3} = 4.0$, $J_{2a,2e} = 12.5$ Hz, H2e); 3.95 (m, $J_{3,4a} = J_{3,4e} = 3.8$, $J_{2e,3} = 4.0$, $J_{2a,3} = 5.2$ Hz, H3); 4.80 (m, $J_{4e,5} = 2.9$, $J_{5,6} = 6.1$, $J_{5,6'} = 6.7$, $J_{4a,5} = 11.7$ Hz, H5).
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 22. ^1H -Nmr (CDCl_3) of **3**: δ 1.87 (ddd, $J_{6R,7} = 4.0$, $J_{5,6R} = 6.9$, $J_{6R,6S} = 14.7$ Hz, H6R); 2.05 and 2.08 (2 s, Ac); 2.18 (ddd, $J_{5,6S} = 6.5$, $J_{6S,7} = 9.1$, $J_{6R,6S} = 14.7$ Hz, H6S); 2.31 (m, $J_{2,4a} = J_{3,4a} = 2.6$, $J_{4a,5} = 11.5$, $J_{4a,4e} = 18.5$ Hz, H4a); 2.49 (m, $J_{2,4e} = 1.2$, $J_{4e,5} = 4.0$, $J_{3,4e} = 6.1$, $J_{4a,4e} = 18.5$ Hz, H4e); 4.07 (m, H10, H10'); 4.49 (m, $J_{4e,5} = 4.0$, $J_{5,6S} = 6.5$, $J_{5,6R} = 6.9$, $J_{4a,5} = 11.5$ Hz, H5); 5.10 (m, H7); 6.02 (ddd, $J_{2,4e} = 1.2$, $J_{2,4a} = 2.5$, $J_{2,3} = 10.0$ Hz, H2) and 6.87 (ddd, $J_{3,4a} = 2.5$, $J_{3,4e} = 6.1$, $J_{2,3} = 10.0$ Hz, H3).
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